G-Biosciences ♦ 1-800-628-7730 ♦ 1-314-991-6034 ♦ technical@GBiosciences.com

A Geno Technology, Inc. (USA) brand name

SpinOUT™ for PCR

(Cat. # 786-174, 786-175)

INTRODUCTION

SpinOUT™ columns are used for the rapid purification and buffer exchange of protein and/or nucleic acid samples**.** Simply apply the samples on top of the column and spin briefly to collect clean samples. These columns are suitable for removing salt, unincorporated radioisotopes, dye, primer, dNTP mix, and buffer exchange. The SpinOUT™ columns are for processing up to 0.1 ml samples.

SpinOUT™-PCR is for cleaning PCR products:

I. **SpinOUT™-PCR20** is for purifying >100bp PCR products from <20bp primers, dyes, unincorporated nucleotides, and salts.

II. **SpinOUT™-PCR32** is for purifying >200bp PCR products from <32bp primers, dyes, unincorporated nucleotides, and salts.

ITEM(S) SUPPLIED

Description	**SpinOUT™-PCR20 Cat. # 786-174**	**SpinOUT™-PCR32 Cat. # 786-175**
SpinOUT™ Columns	10 columns	10 columns

STORAGE CONDITIONS

It is shipped at ambient temperature. Upon arrival, store at 4°C. When stored and used properly, the columns are good for 1 year.

ADDITIONAL ITEMS REQUIRED

Collection tubes

SPECIFICATIONS

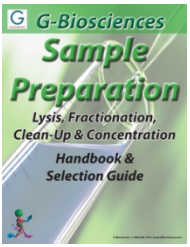
	SpinOUT™-PCR20	**SpinOUT™-PCR32**
Sample Volume	20-100µl	
Storage Buffer	TE Buffer (10mM Tris-HCl, 1mM EDTA, pH7.6	
Average Particle Size	50µm	
pH Stability Working Range	3-11	
DNA Exclusion Limit	118bp	271bp
Globular Protein Exclusion Limit	~2 x 10^6 (2,000kDa)	~9 x 10^6 (9,000kDa)

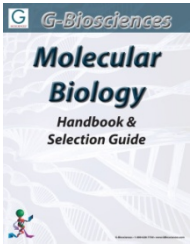
PROTOCOL

1. Invert the column several times to resuspend the gel material. Spin the column for 10 seconds at 100 x g to allow the gel to collect in the column
   NOTE: *Do not spin harder than recommended or damage to resin may occur.*
2. Remove the tip of the column and let the liquid drain into a collection tube.
3. Buffer Equilibration**:** SpinOUT™ PCR columns are in 20% ethanol. Equilibrate the column with a buffer of your choice.
4. Apply about 0.1-0.2ml of desired buffer into the columns and let the buffer drain into the collection tube. Repeat this process 3 times, and discard the liquid collected in the collection tube.
5. Place the column in a 2ml centrifuge tube. Centrifuge at 1000xg for 2 minutes, then discard the liquid collected in the centrifuge tube.
6. Place the column back in the same centrifuge tube. Carefully apply sample (20-100µl) to the center of the column without disturbing the resin bed. Wait for 1-2 minutes.
   NOTE: *The maximum volume you should load is 100µl.*
7. After loading the column, place the column in a new and clean collection tube and centrifuge at 1000xg for 2 minutes. Collect the liquid containing purified sample.
8. Discard used column.

RELATED PRODUCTS

Download our Sample Preparation and Molecular Biology Handbooks.





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