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A Geno Technology, Inc. (USA) brand name

RUBEO™

A Fluorescent Protein Gel Stain

(Cat. # 786-644, 786-645)

INTRODUCTION

RUBEO™ is a fluorescent protein stain that is ideal for staining proteins in 1D and 2D electrophoresis gels. The fluorescent stain has a red emission at 605nm. The fluorescent stain has nanogram sensitivity and is compatible with mass spectroscopy, microsequencing and immunostaining. The stained gels can be visualized on a simple UV transilluminatoror the most complex laser scanners.

ITEMS INCLUDED

Description	Cat. # 786-644	Cat. # 786-645
RUBEO™	200ml	1L

STORAGE CONDITION

Shipped at ambient temperature. On arrival store at 4°C. When stored and used properly RUBEO™ is stable for one year.

ADDITIONAL ITEMS REQUIRED

- G-Biosciences' SDS-PAGE Gel Fixing Solution (Cat. # 786-236) or 30% ethanol and 10% acetic acid
- G-Biosciences' Destain I (Cat. # 786-526) or 40% ethanol and 10% acetic acid
- Deionized water

IMPORTANT NOTES

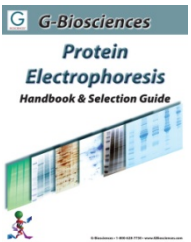
- Gel clarity will depend greatly on the quality and purity of the reagents used in making and running of the gel as well as the quality of the protein sample loaded on the gel.
- Always use clean containers and highly purified deionized water for fixing and staining the gel. All % are in V/V.
- Never touch the gel with fingers. **Wear gloves during all gel handling.**
- The volumes used are for mini gels, adjust accordingly for larger protein gels.

PROTOCOL

1. Allow the reagents to reach room temperature.
2. Following electrophoresis, fix the gel in 100ml 30% ethanol and 10% acetic acid. For isoelectric focusing (IEF) gels, fix the gel first in 20% TCA for 30 minutes. Fix the gels for 30 minutes to 3 hours depending on gel size and thickness, 30 minutes is suitable for mini gels. Gels can be fixed overnight with no detrimental effects.
3. Rinse the gel in 50ml 20% ethanol. Rinse 3 x 10 minutes
4. Add 50ml RUBEO™ stain to the rinsed gel. Wrap the staining container in aluminum foil to protect from direct light. Stain the gel for six hours with gentle shaking.
5. Equilibrate the gel in 50ml deionized water by rinsing 2 x 10 minutes.
   NOTE: *A scan can be taken at this stage, however a high background will be present. For optimal staining continue to the next step.*
6. Destain the gel 100ml 40% ethanol and 10% acetic acid overnight with gentle shaking.
7. Equilibrate the gel in 50ml deionized water by rinsing 2 x 10 minutes.
8. Visualize the gel on a UV transilluminator or scan the gel.

RELATED PRODUCTS

Download our Protein Electrophoresis Handbook.



http://info.gbiosciences.com/complete-protein-electrophoresis-handbook

For other related products, visit our website at www.GBiosciences.com or contact us.

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