



A Geno Technology, Inc. (USA) brand name

Well-Coated[™] Protein A, Protein G & Protein A/G

96-Well Plates Coated with Immunoglobulin Binding Protein A, Protein G or Protein A/G

(Protein A: Cat. # 786-730, 786-731, 786-770, 786-771) (Protein G: Cat. # 786-732, 786-733, 786-774, 786-775) (Protein A/G: Cat. # 786-734, 786-735, 786-772, 786-773)



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INTRODUCTION

Well-Coated^{$^{\infty}$} Protein A, Protein G and Protein A/G plates are designed to bind the constant (F_c) region of immunoglobulins ensuring that the antigen binding domain of the antibody is orientated away from the plate, offering maximum exposure of the binding site. The immunoglobulin orientation on the Well-Coated^{$^{\infty}$} Protein A, Protein G and Protein A/G plates improves the antibody capacity compared to plates that are coated directly with antibodies.

The choice of plate is dependent on the antibody type to be used. See the appendix a table highlighting the binding affinity for Protein A, Protein G and Protein A/G.

Well-Coated™ Protein A, Protein G and Protein A/G plates are for single antibody assays and are not suitable for multiple assays (sandwich ELISAs) as the first antibody will not block all IgG binding sites and therefore false positives will occur with the second antibody. The wells are coated to a 100µl depth and are supplied pre-blocked in our Superior™ Blocking Buffer that contains an antigenically non-determining protein. The plates are protected with our WellCoat™ Stabilizer (Cat. # 786-1217) that creates a protective layer over the immobilized agents. The reagent will not interfere with the assay and has no effect of the efficiency or capacity of the wells. The WellCoat™ Stabilizer offers greater protection and shelf life of the plates. In some cases, the protective layer may give the appearance of a white coating. The clear, white and black plates are offered for colorimetric, chemiluminescence and fluorescent detection systems, respectively.

ITEM(S) SUPPLIED

Cat. #	Components	Size
786-730	Well-Coated™ Protein A Coated 96 well plate	5 plates
786-731	Well-Coated™ Protein A Coated 8-well strip plate	5 plates
786-770	Well-Coated™ Protein A Coated 96 well plate, Black	5 plates
786-771	Well-Coated™ Protein A Coated 96 well plate, White	5 plates
786-732	Well-Coated™ Protein G Coated 96 well plate	5 plates
786-733	Well-Coated™ Protein G Coated 8-well strip plate	5 plates
786-774	Well-Coated™ Protein G Coated 96 well plate, Black	5 plates
786-775	Well-Coated™ Protein G Coated 96 well plate, White	5 plates
786-734	Well-Coated™ Protein A/G Coated 96 well plate	5 plates
786-735	Well-Coated™ Protein A/G Coated 8-well strip plate	5 plates
786-772	Well-Coated™ Protein A/G Coated 96 well plate, Black	5 plates
786-773	Well-Coated™ Protein A/G Coated 96 well plate, White	5 plates

STORAGE CONDITIONS

Shipped at ambient temperature. Upon arrival, store unopened at 4°C. Once opened the plates can be stored in a resealable bag (ZipLoc) with an appropriate desiccant at 4°C.

BINDING CAPACITY

Well-Coated[™] Protein A: ~4pmol rabbit IgG/well
Well-Coated[™] Protein G: ~2pmol rabbit IgG/well
Well-Coated[™] Protein A/G: ~5pmol rabbit IgG/well

BASIC ELISA ASSAY PROTOCOL

The following protocol is a simple ELISA protocol and the protocol and reagents used will have to be optimized for specific applications and assays.

Items Needed But Not Supplied

- Antibody to be bound to plate (see Appendix for correct Well-Coated[™] plate to be used).
- Wash Buffer: femtoTBST™ (Cat. # 786-161) or femtoPBST™ (Cat. # 786-162); 10X concentrated wash buffers supplemented with Tween® 20. Or an appropriate wash buffer of choice.
- Blocking Buffer: A suitable blocking buffer, we recommend our Superior™ Blocking Buffer (Cat. # 786-655 to 786-661) or NAP-BLOCKER™, an animal free blocking agent suitable for ELISA (Cat. # 786-190).
- Labeled Antigen, visit <u>www.GBiosciences.com</u> for horseradish peroxidase (HRP), alkaline phosphatase (AP) and biotin labeling kits.
- Detection system, femtoELISA™ is a chromogenic detection system for HRP and AP (Cat. # 786-110 to 786-113)

Protocol

- 1. Wash the wells to be used three times with 300µl Wash Buffer.
- Dilute the antibody to be bound to ~1µg/ml with the Blocking Buffer. Add up to 100ul to each well.
- Incubate at room temperature for 30-60 minutes, for optimal binding use a plate shaker.
- 4. Wash each well three times with 300µl Wash Buffer.
- Add the labeled antigen at a concentration of ~0.1µg/well, diluted in Blocking Buffer, if necessary.
- Incubate at 37°C for 1 hour.
- 7. Wash each well three times with 300µl Wash Buffer.
- 8. Detect the label signal according to the manufacturer's instructions, using 200µl detection reagent per well.

NOTE: For biotin, incubate the plate for a further 1 hour at 37°C with an enzymelabeled streptavidin or other biotin detection system. Wash as before and then detect the signal.

APPENDIX

Species	Antibody Class	Protein A	Protein G	Protein A/G
Mouse	Total IgG	****	****	****
	IgM	-	-	-
	lgG1	*	***	***
	lgG2a	****	****	****
	lgG2b	****	****	****
	IgG3	****	****	****
	Total IgG	*	***	***
	lgG1	*	***	***
Rat	lgG2a	-	****	****
	lgG2b	-	*	*
	lgG2c	****	****	****
	Total IgG	****	****	****
	lgG1	****	****	****
	lgG2	****	****	****
	IgG3	*	****	****
Human	IgG4	****	****	****
nulliali	IgM	*	-	*
	IgD	-	-	-
	IgA	*	-	*
	Fab	*	*	*
	ScFv	*	-	*
	Total IgG	*	****	****
Goat	lgG1	*	****	****
	lgG2	****	****	****
Dog	Total IgG	****	*	****
Guinea Pig	Total IgG	****	*	****
Pig	Total IgG	****	*	****
Rabbit	Total IgG	****	****	****
	Total IgG	*	****	****
Sheep	lgG1	*	****	****
	lgG2	****	****	****

Cat	Total IgG	****	*	****
Chicken	Total IgY	-	-	-
Cow	Total IgG	*	****	****
	lgG1	*	****	****
	lgG2	****	****	****
Horse	Total IgG	*	****	****
	IgG(ab)	*	-	*
	IgG(c)	*	-	*
	IgG(T)	-	****	****

Table 1: Relative affinity of Protein A, Protein G and Protein A/G for Immunoglobulins

RELATED PRODUCTS

Download our Sample Preparation Handbook



http://info.gbiosciences.com/complete-protein-sample-preparation-handbook/

For other related products, visit our website at <u>www.GBiosciences.com</u> or contact us._{Last} saved: 9/7/2017 CMH

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